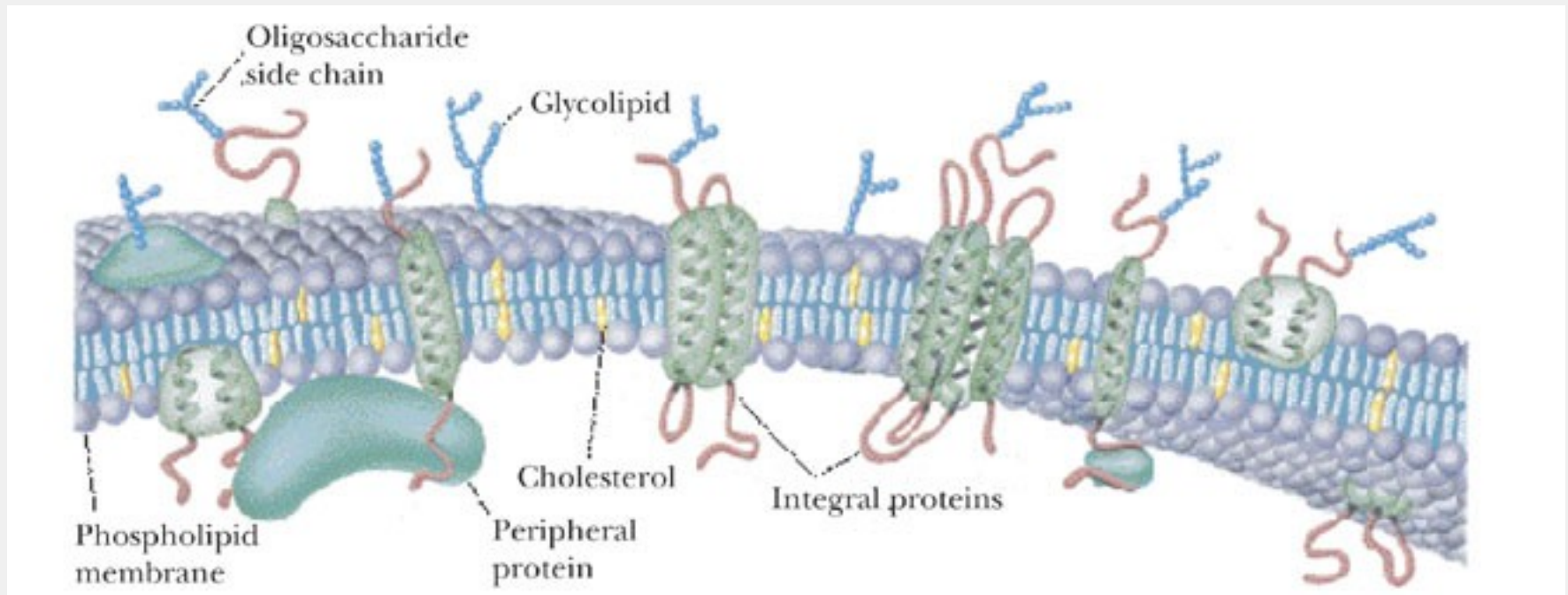


# Chapter 12: Membranes

**Voet & Voet:  
Pages 390-415**

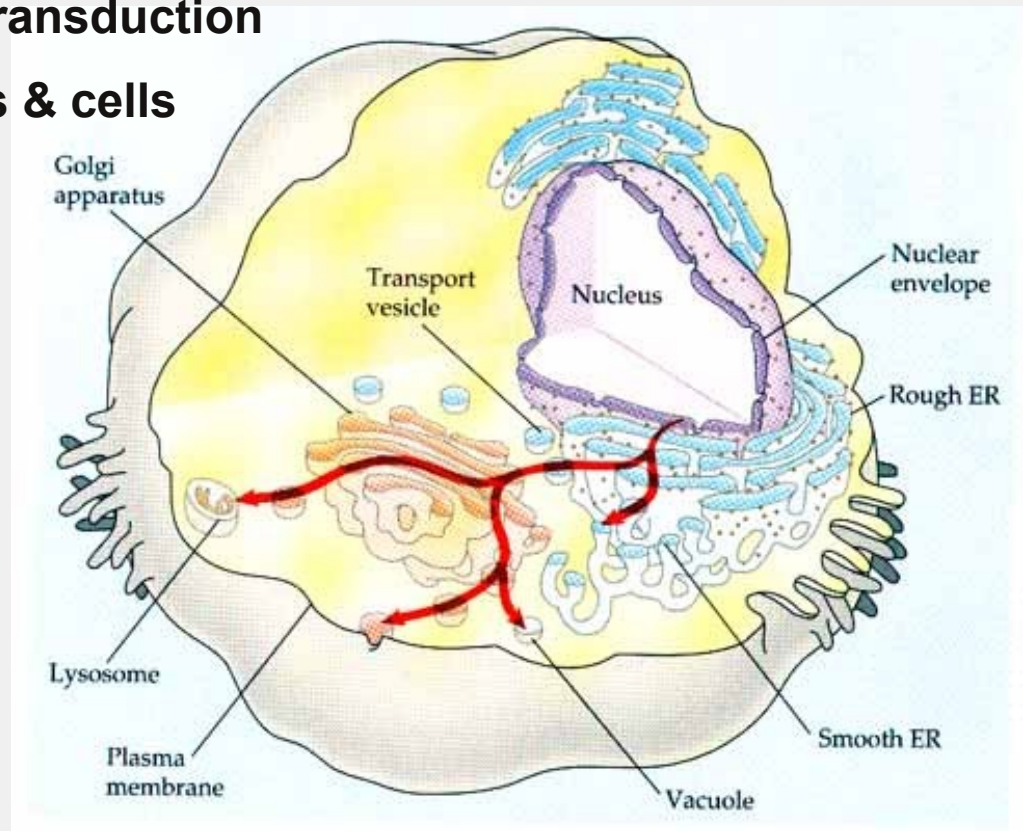


# Membranes

## Essential components of all living cells (define boundry of cells)

- exclude toxic ions and compounds; accumulation of nutrients
- energy transduction; cell locomotion
- reproductive processes; signal transduction
- varied interaction with molecules & cells

Some of the membranes  
present in a eucaryotic cell



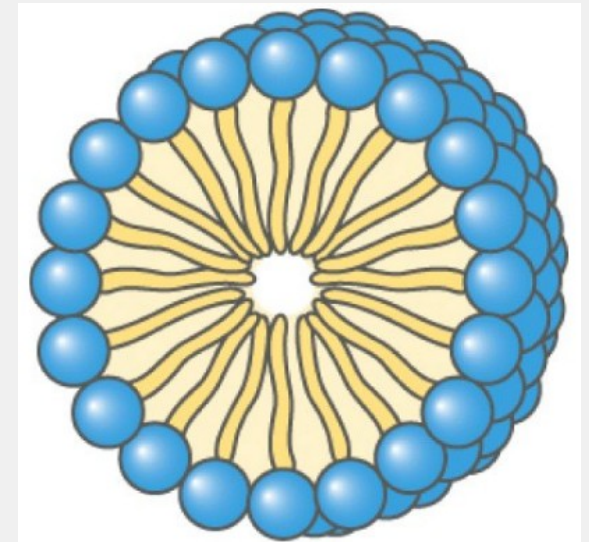
# Simple Lipid Monolayers

Single Tailed lipids form **micelles** in H<sub>2</sub>O

- globular (spherical) aggregates whose hydrocarbons tails are out of contact with water (similar to soaps and detergents)
- Eliminates unfavorable contacts between water and the hydrophobic tails AND permits solvation of polar head groups

Micelle formation is a cooperative process

- an assembly of a just a few lipids cannot shield their tails from the solvent
- in dilute solutions lipids do not form micelles until their concentration surpasses a **critical micelle concentration** (*cmc*)
- above the cmc almost all added lipids aggregate to form micelles

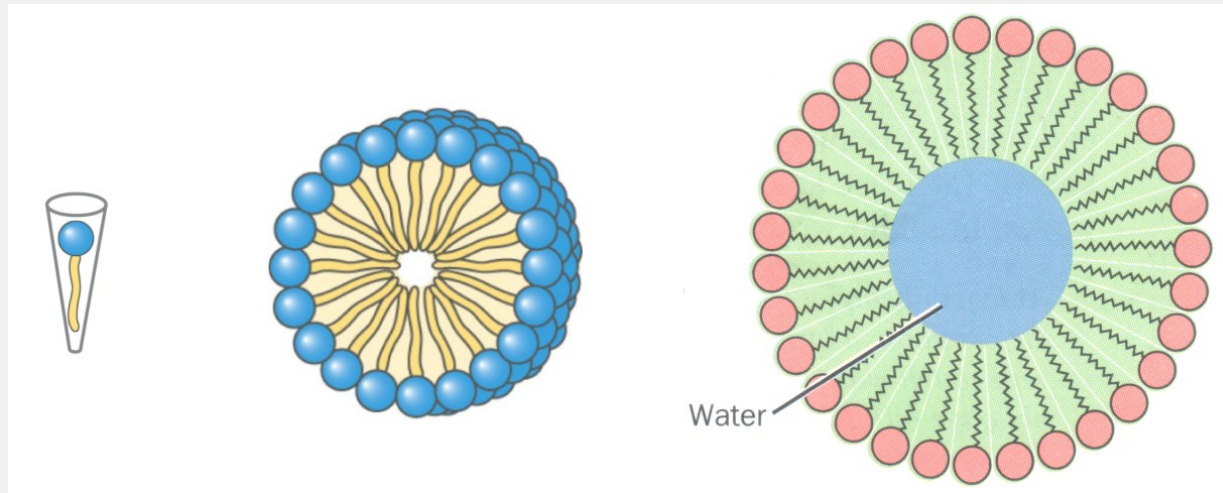


Micelle composed of single tailed lipids

# Micelle Size & Shape

## Geometric shape of lipid dictates optimal size & shape

- single tailed lipids have a conical van der Waal's surface
- efficiently packs into a spheroidal micelle with an ~ constant number of molecules (typically several hundred)
- larger micelles would have an internal water filled cavity (energetically unfavorable)



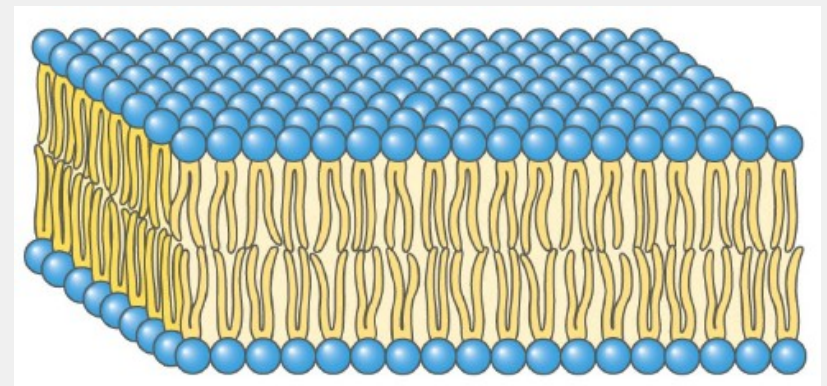
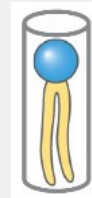
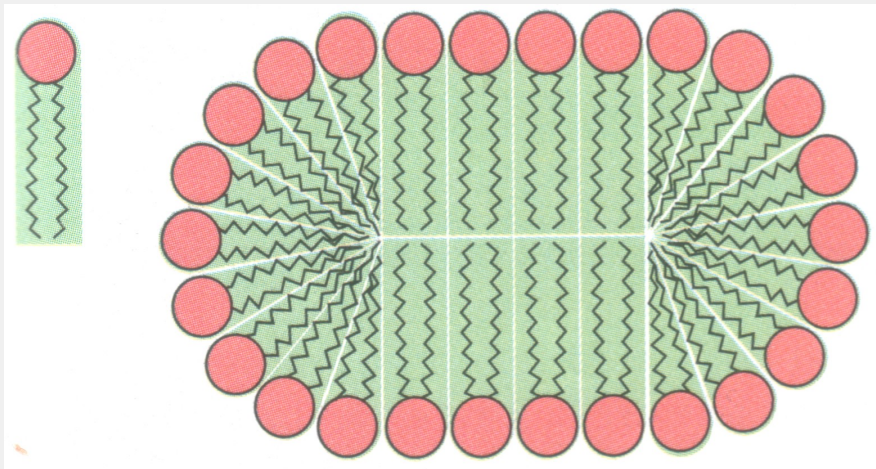
Typical Micelle

Energetically Unfavorable  
(does not occur normally)

# Simple Bilayers

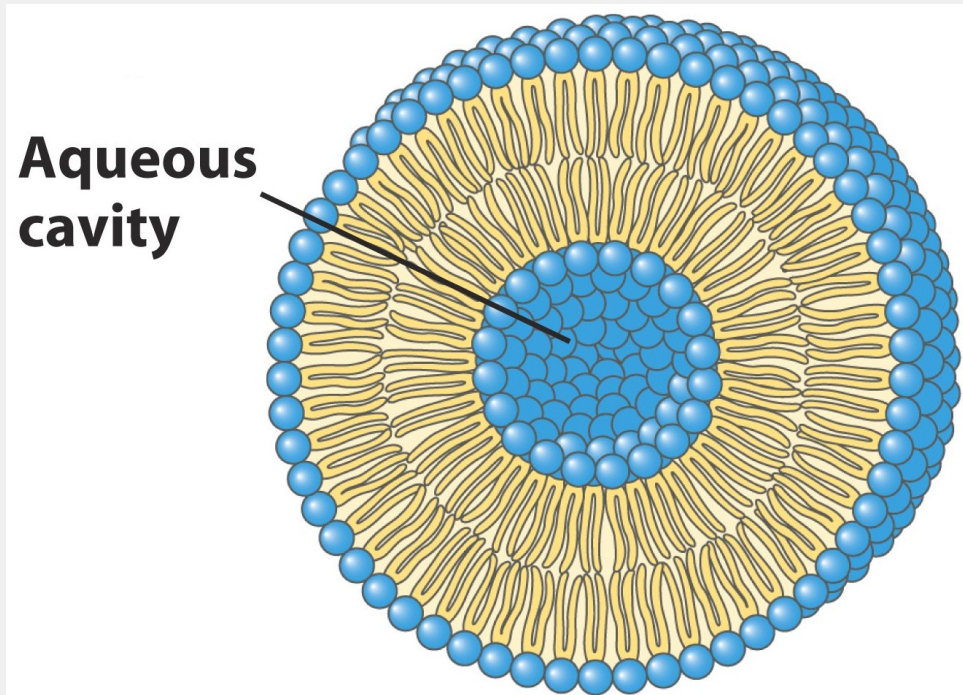
## Glycerophospholipids and Sphingolipids are two tailed lipids

- two tailed lipids have a roughly cylindrical van der Waal's surface and preferentially form disc-shaped micelles
- Physical studies (Electron Microscopy & Small Angle X-ray Scattering) indicate the hydrophobic tails are almost fully extended
  - 60 Å cross section – length of one lipid ~30 Å when fully extended





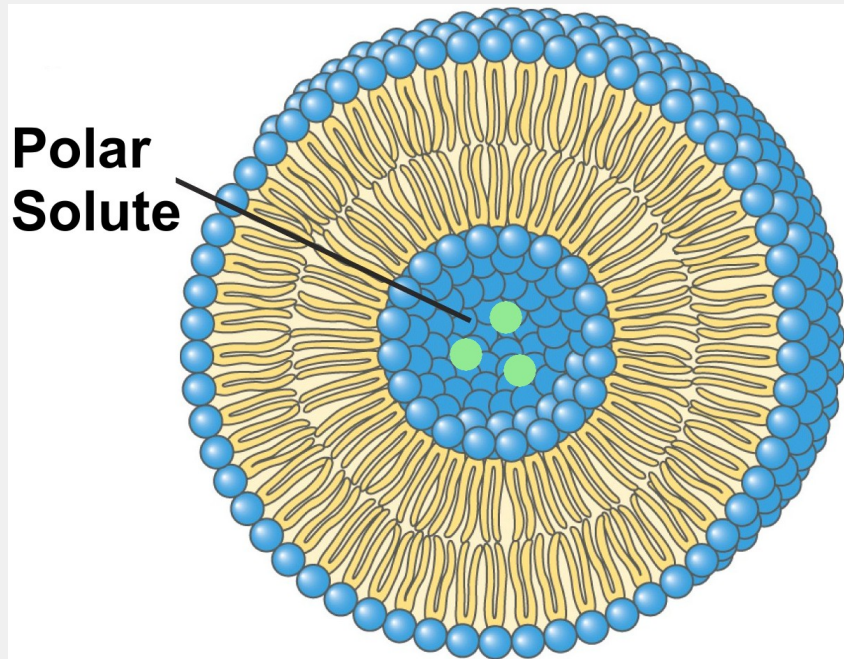
# Liposomes



## Phospholipid suspensions in water form multilamellar vesicles

- sonication generates liposomes
- generally from 100's – 1000 Å in diameter
  - central cavity filled with solvent similar to biological membranes
- once formed, liposomes are stable and can be isolated
  - important tool for studying membrane behavior

# Liposomes



**Lipid bilayers are impermeable to most polar substance**

- **sonication in presence of polar substance forms liposomes with polar compound in interior**
  - **polar compounds do not escape when liposomes are dialysed**
- **(surprisingly) water can traverse bilayers**

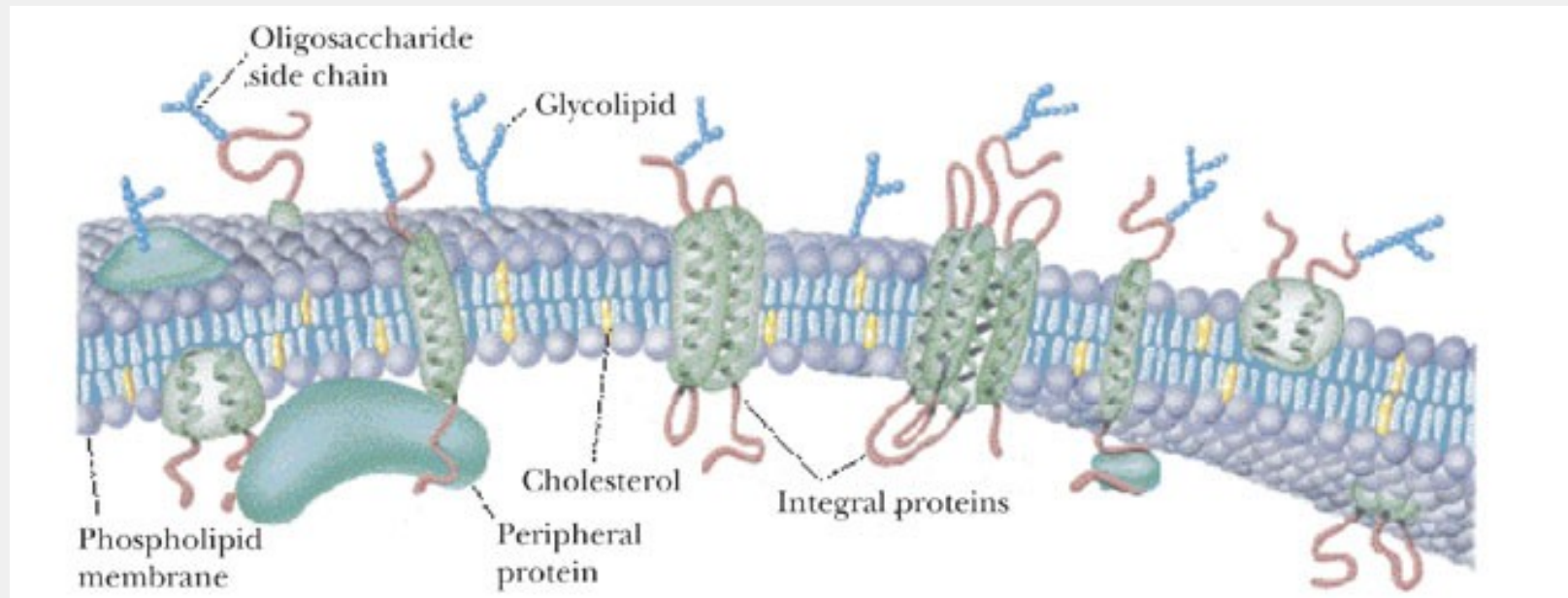
**Liposomes are a promising vehicle for “drug” delivery**

- **requires ability to target vesicles to cells**

# Fluid Mosaic Model of Membranes

Singer & Nichol森 propose **Fluid Mosaic** Model of Membranes

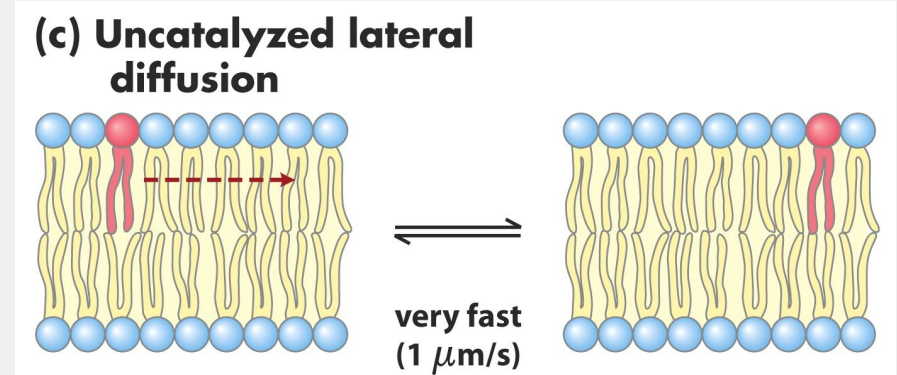
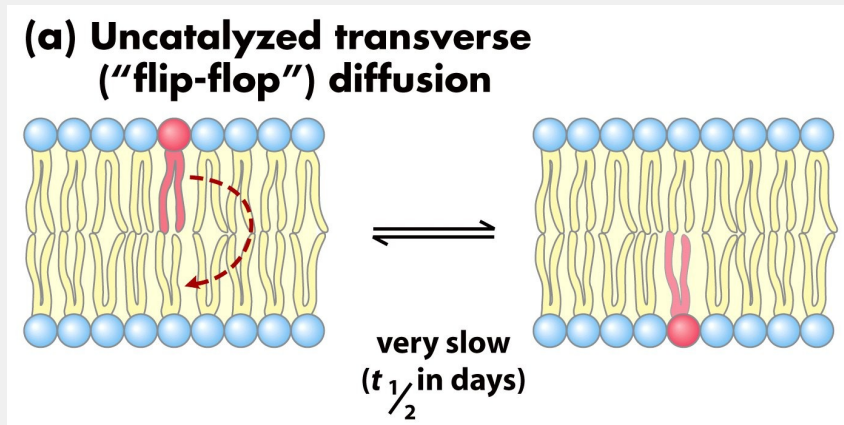
- membranes are dynamic structures
- lipids and proteins are free to move in 2D



# Lipid Bilayer

Bilayers are best described as two-dimensional fluids

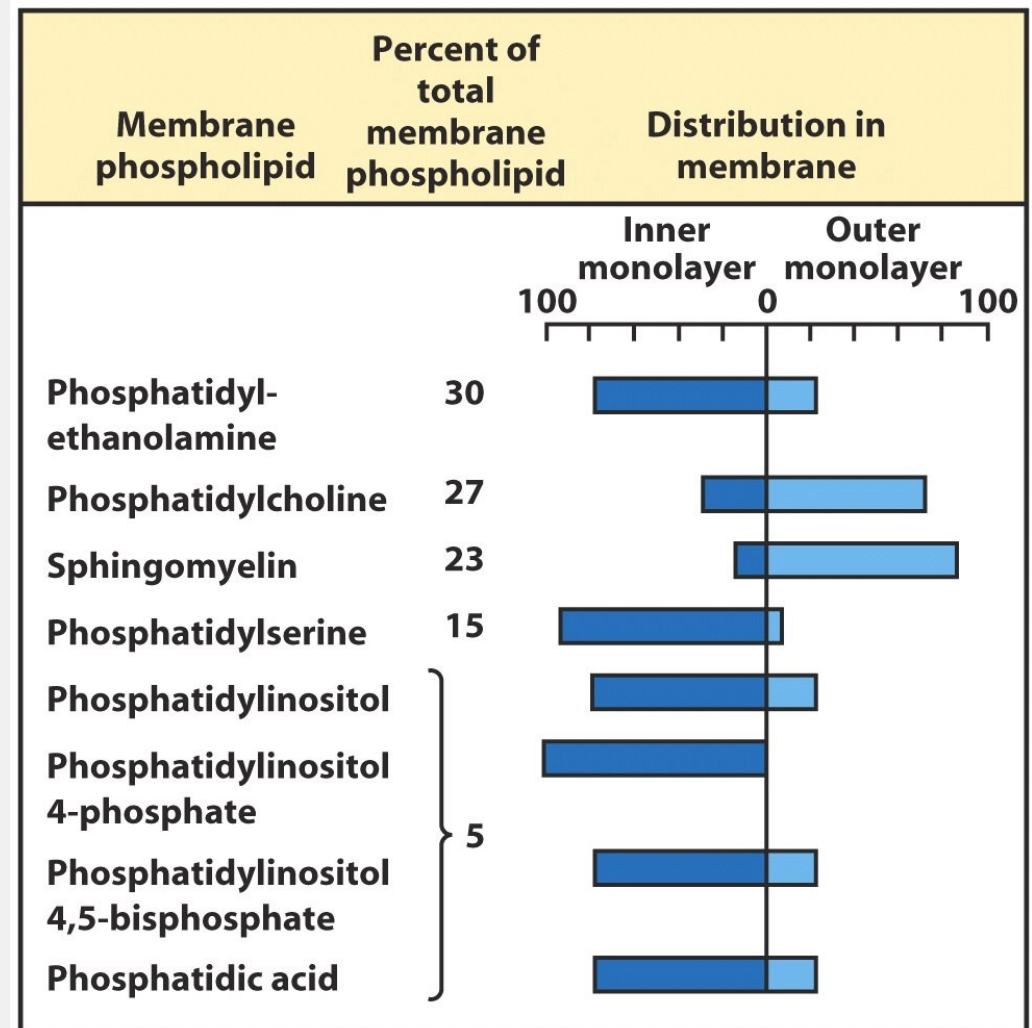
- lipid transfer across the membrane is rare (transverse diffusion or 'flip-flop')
- lipids are highly mobile in the plane of the bilayer (lateral diffusion)



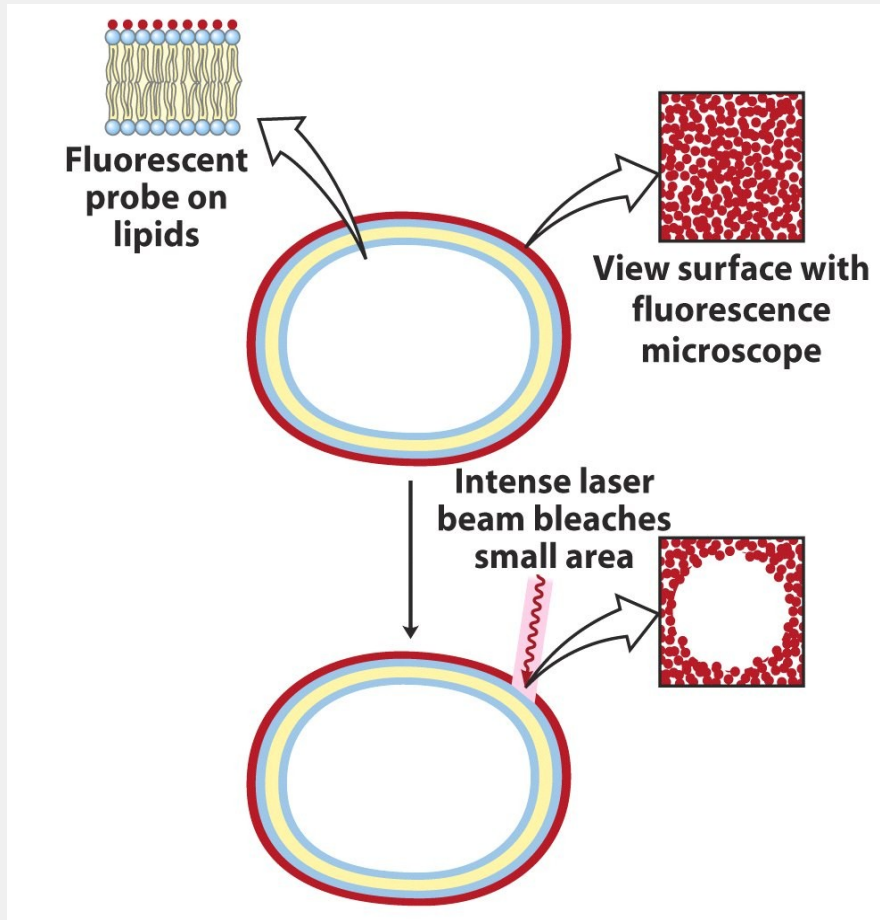
# Asymmetric Lipid Distributions

In biological membranes, the slow rate of transverse diffusion contributes to the asymmetric distribution of lipids

- consequently, the inner and outer membrane monolayers have different properties

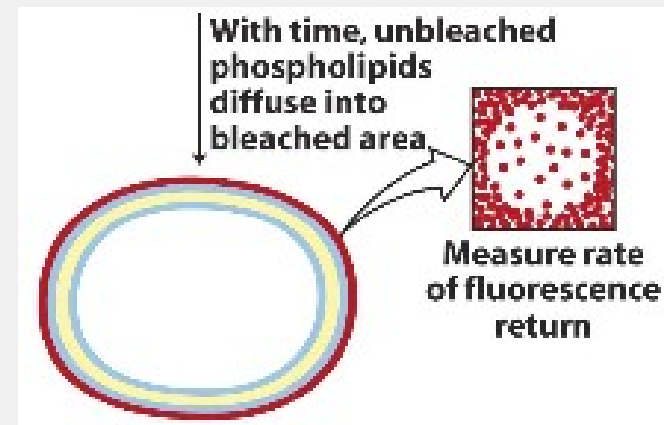


# Evidence for Lateral Diffusion



## Photobleaching Experiment

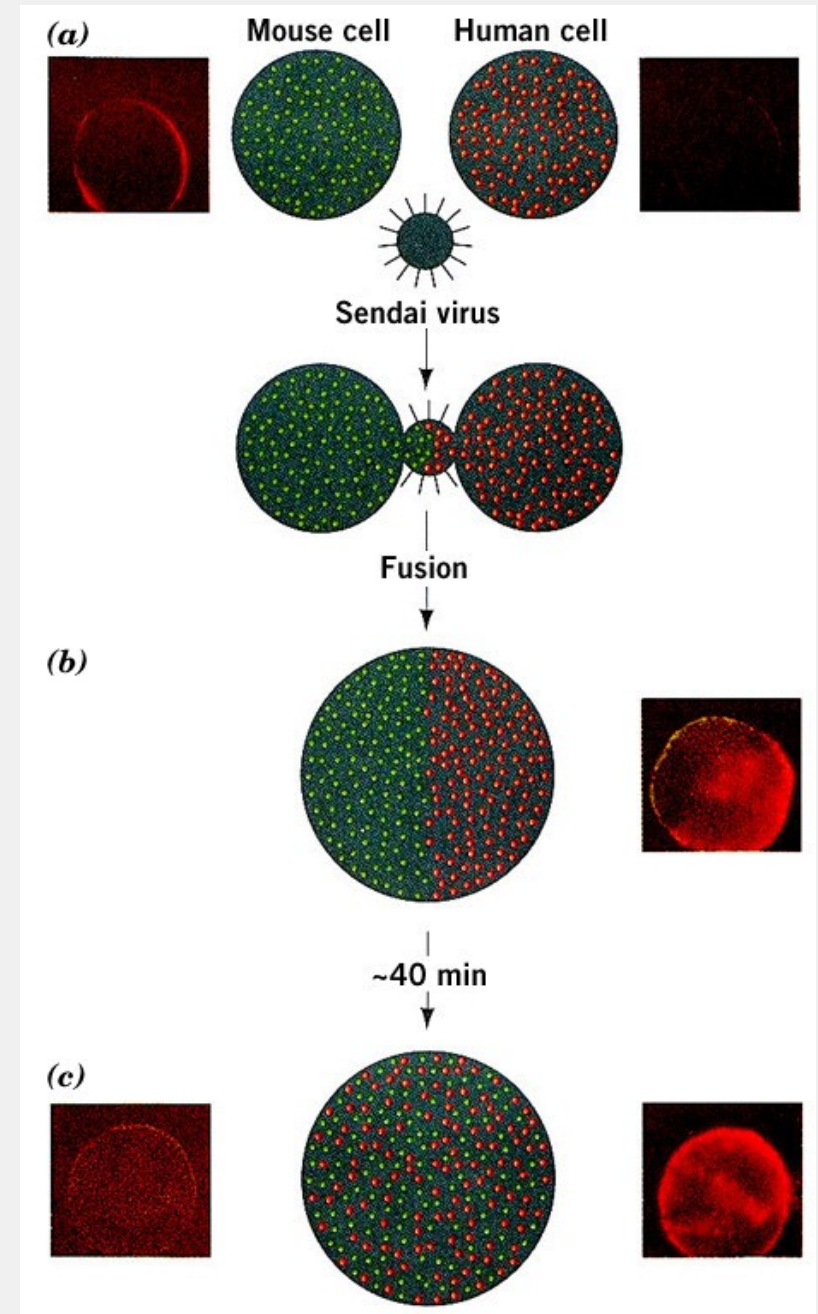
- Can observe lateral diffusion of labeled lipids as a function of time



# More Evidence

## A second experiment showing lateral diffusion in bilayers

- separate cells are labeled with distinct fluorophores
- cell membranes are fused
- fluorophore movement is observed as function of time



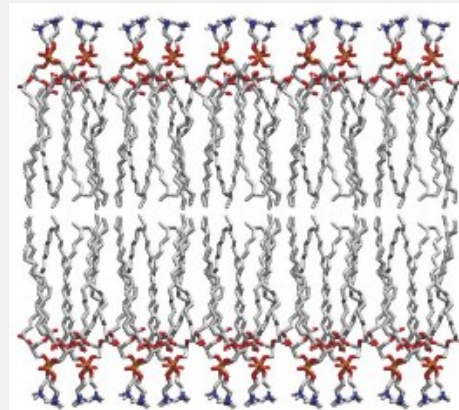
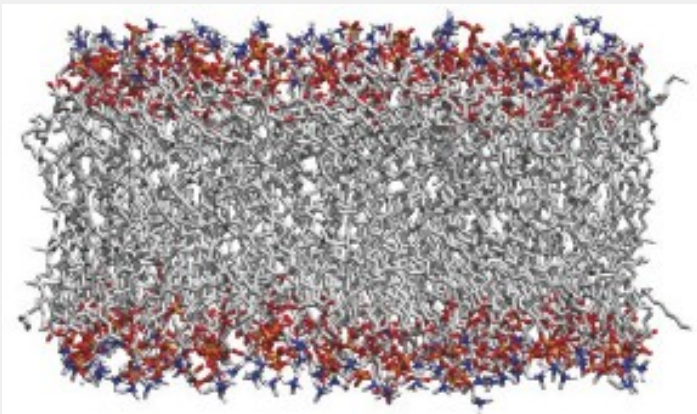
# Bilayers and Temperature

Below a characteristic transition temperature, bilayer undergo a sort of phase change called the **order-disorder transition**

- below the transition temperature, bilayers become a gel-like solid and lose their fluidity
  - average spacing between hydrocarbon tails is reduced from 4.6 to 4.2 Å
- transition temperature increases with longer tail lengths and greater saturation (typically between 10 – 40° C)

eg. longer tails and more saturation favor ordered state

Cholesterol decreases transition temperature by inhibiting formation of the gel like state



Disordered or fluid (left)  
and Ordered (right)  
states of a bilayer

# Bilayers and Temperature

The composition of bilayers changes as a function of temperature of growth

- increasing fraction longer tailed and saturated lipids at higher temperature

eg. 22% of lipid is saturated at 10 °C (4/22 or 18% is 14:0)  
56% of lipid is saturated at 40 °C (8/56 or 14% is 14:0)

## *E. coli* culture temperature

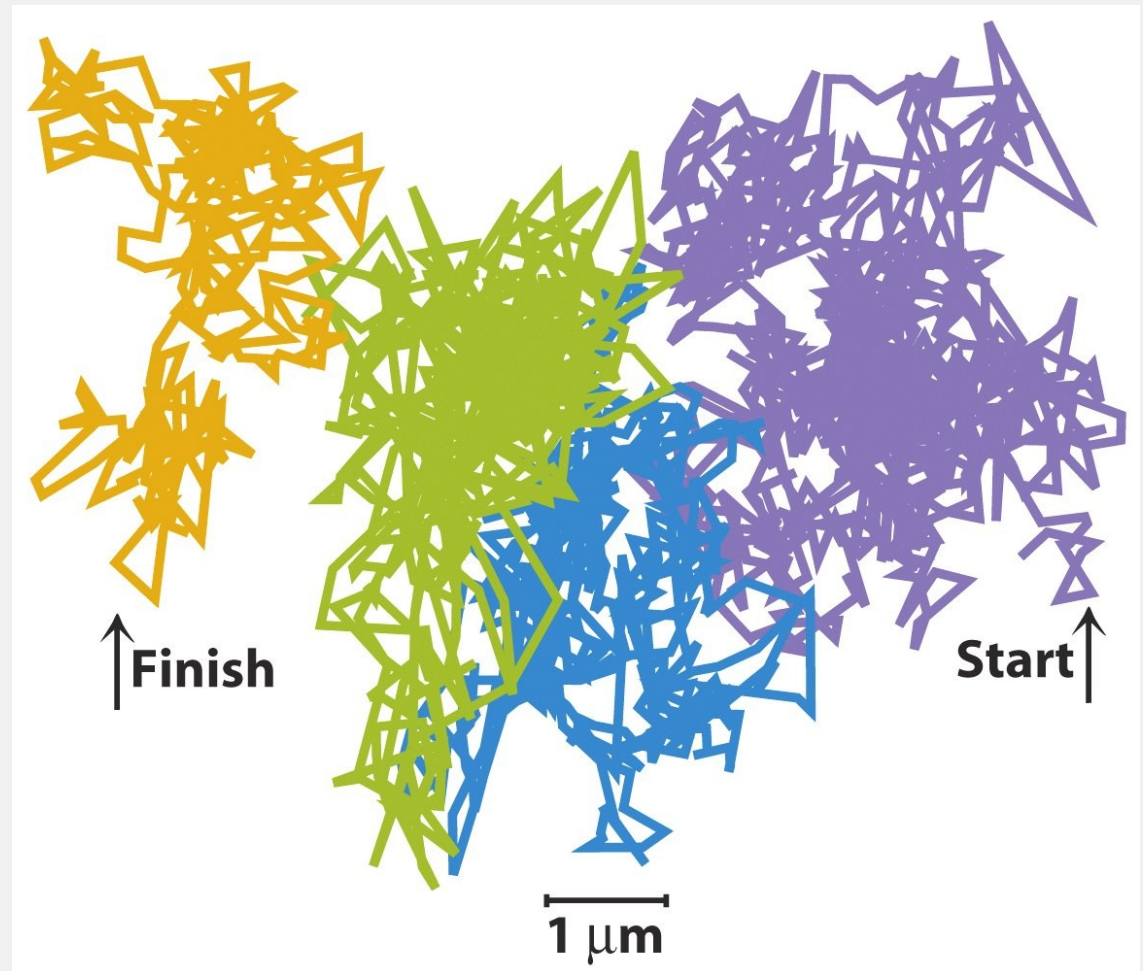
	Percentage of total fatty acids*			
	10 °C	20 °C	30 °C	40 °C
Myristic acid (14:0)	4	4	4	8
Palmitic acid (16:0)	18	25	29	48
Palmitoleic acid (16:1)	26	24	23	9
Oleic acid (18:1)	38	34	30	12
Hydroxymyristic acid	13	10	10	8
Ratio of unsaturated to saturated <sup>†</sup>	2.9	2.0	1.6	0.38

# Explain this Result

**The movement of a single labeled lipid on the cell surface**

- recorded on video over 56 ms (fluorescent microscopy)
- starts in the purple area then moves through the blue, green and final the orange areas.

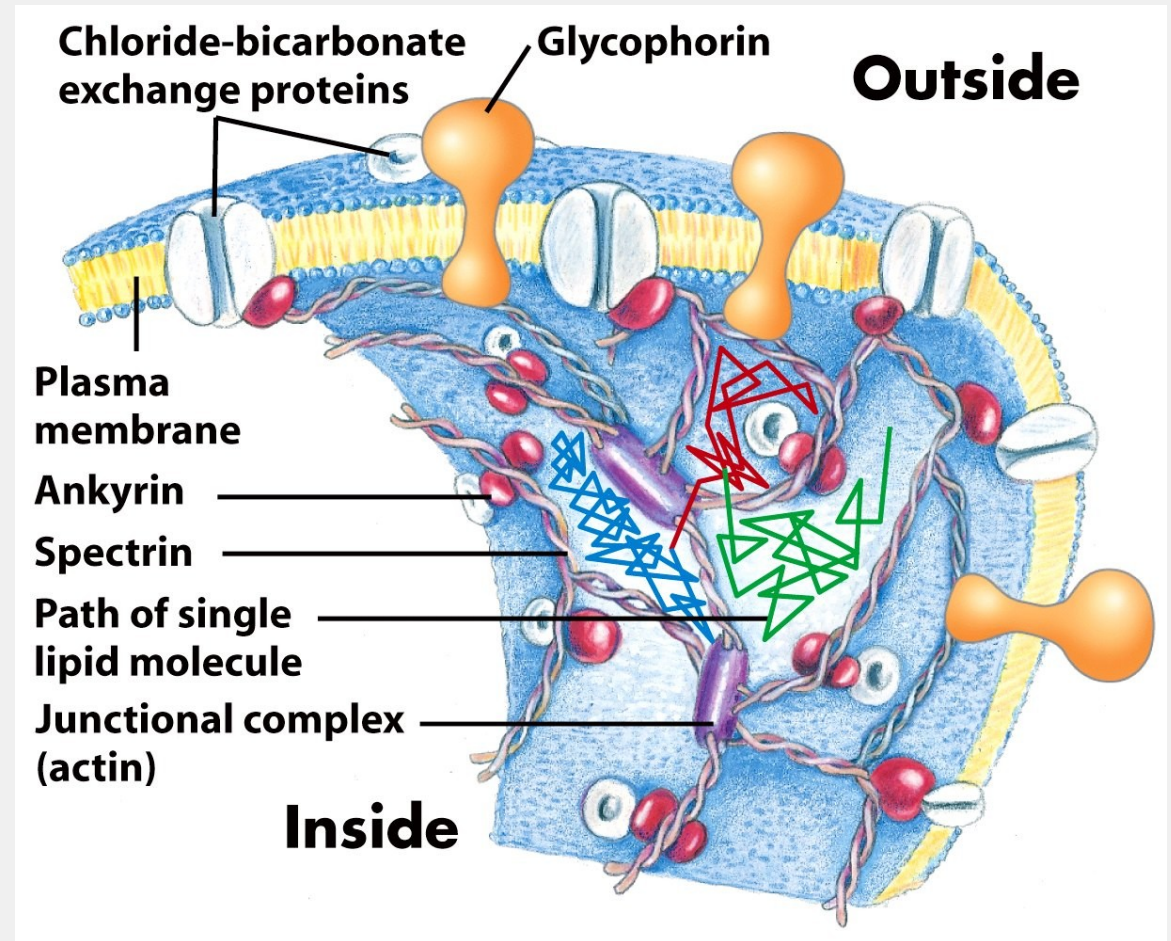
**What's going on?**



# Membranes contain Proteins

The movement is restricted by membrane proteins

- Biological membranes contain significant quantities of protein
- Typical membranes have a protein:lipid ratio around 1:1



# Membrane Composition (eg.)

Biological membranes contain a lot of proteins

Protein:Lipid ratio varies between cell types

**TABLE 12-4** Compositions of Some Biological Membranes

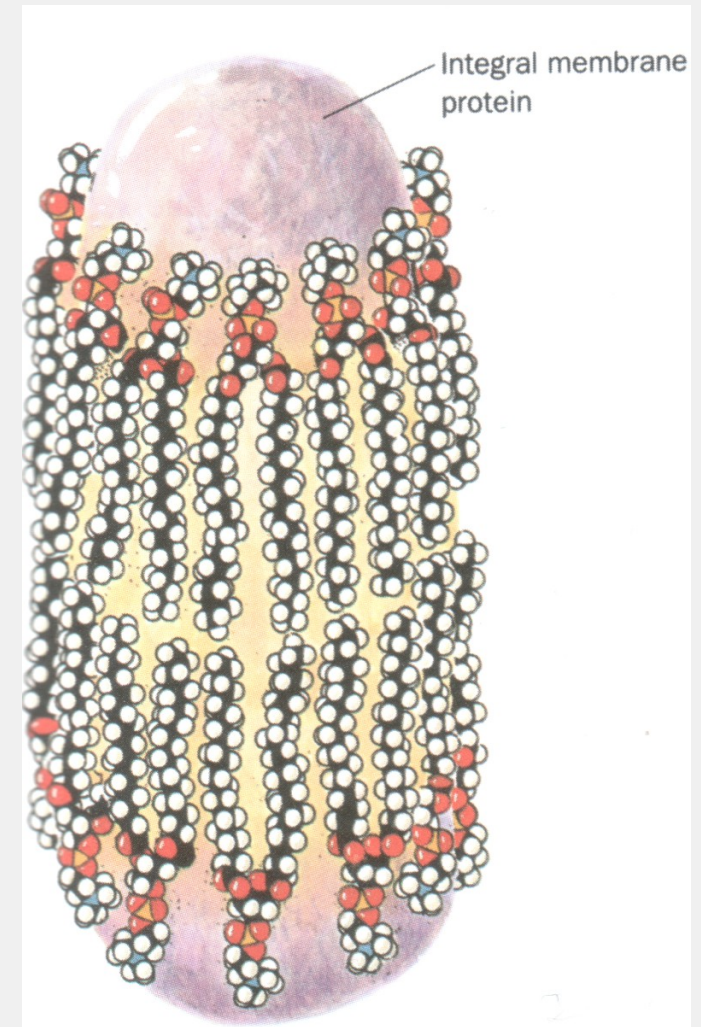
Membrane	Protein (%)	Lipid (%)	Carbohydrate (%)	Protein to Lipid Ratio
Plasma membranes:				
Mouse liver cells	46	54	2-4	0.85
Human erythrocyte	49	43	8	1.1
Amoeba	52	42	4	1.3
Rat liver nuclear membrane	59	35	2.0	1.6
Mitochondrial outer membrane	52	48	(2-4) <sup>a</sup>	1.1
Mitochondrial inner membrane	76	24	(1-2) <sup>a</sup>	3.2
Myelin	18	79	3	0.23
Gram-positive bacteria	75	25	(10) <sup>a</sup>	3.0
<i>Halobacterium</i> purple membrane	75	25		3.0

<sup>a</sup>Deduced from the analyses.

# Membrane Proteins

## Two general classes of membrane proteins

- (1) **Integral** or intrinsic membrane proteins are tightly bound by hydrophobic forces
  - must disrupt membrane (*ie.* detergents) to separate proteins
- (2) **Peripheral** or extrinsic membrane proteins are anchored or associate with membranes
  - can be separated by mild treatments that leave membranes intact

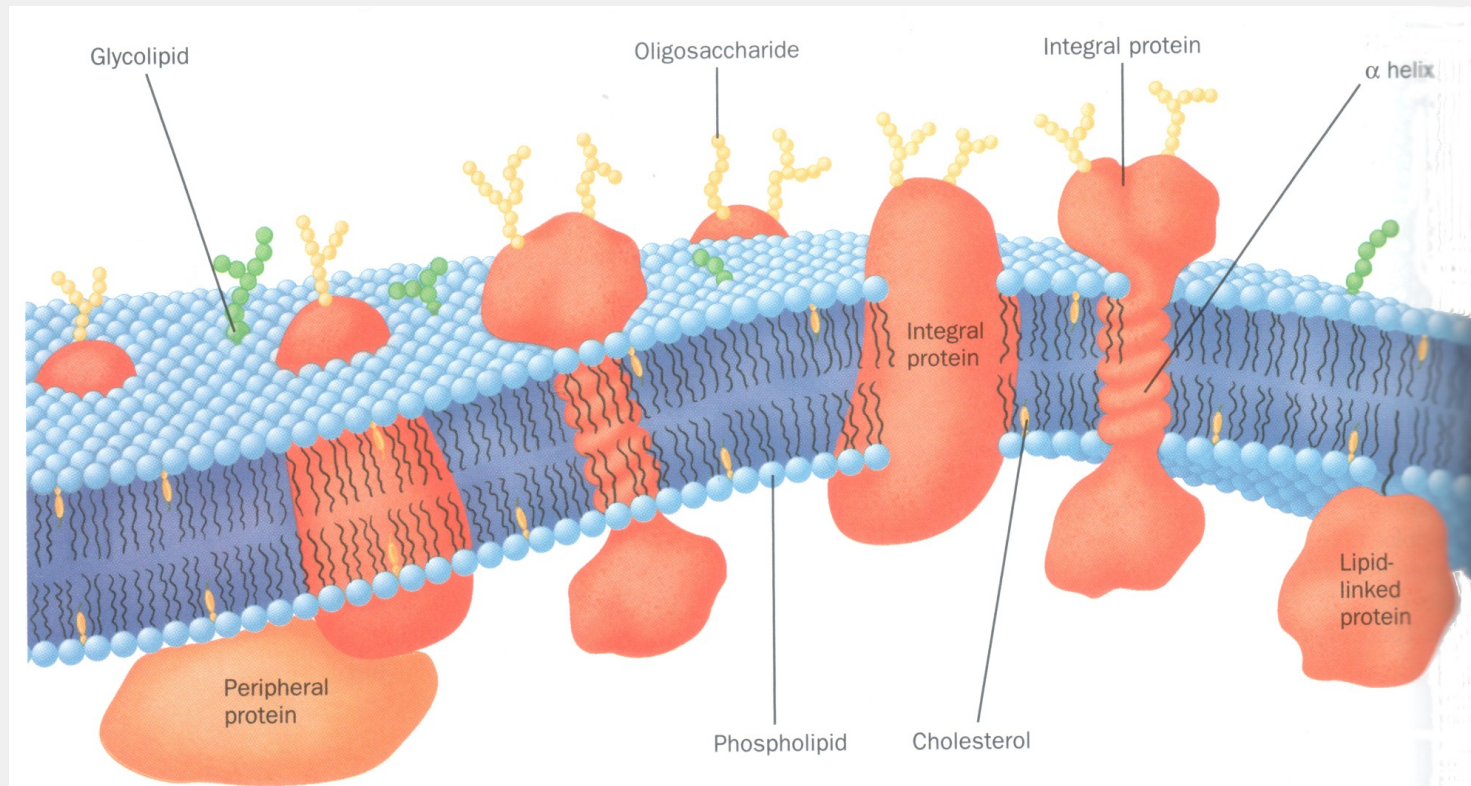


# Membrane Proteins (more)

Integral membrane proteins are asymmetrically oriented

- surface labeling identifies two distinct types of integral proteins

(1) **transmembrane** proteins that span the membrane and are exposed on both sides of the membrane



# Membrane Proteins (cont.)

(2) **embedded** proteins that are only exposed on a specific side of the membrane

- anchored to membrane by polypeptide segment OR lipid modification

Lipid modification that anchor embedded proteins:

- (1) isoprenoid groups
- (2) fatty acyl groups
- (3) glycoinositol phospholipids

